

1 **Global diversity and oceanic divergence of humpback whales (*Megaptera***  
2 ***novaeangliae*)**

3

4 **Research Article**

5

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28 **Running Title:** Humpback genomic diversity and population structure

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1 Humpback whales (*Megaptera novaeangliae*) annually undertake the longest migrations  
2 between seasonal feeding and breeding grounds of any mammal. Despite this dispersal  
3 potential, discontinuous seasonal distributions and migratory patterns suggest that  
4 humpbacks form discrete regional populations within each ocean. To better understand  
5 the worldwide population history of humpbacks, and the interplay of this species with the  
6 oceanic environment through geological time, we assembled mitochondrial DNA control  
7 region sequences representing ~2,700 individuals (465bp, 219 haplotypes) and 8 nuclear  
8 intronic sequences, representing ~70 individuals (3,700bp, 140 alleles) from the North  
9 Pacific, North Atlantic and Southern Hemisphere. Bayesian divergence time  
10 reconstructions date the origin of humpback mtDNA lineages to the Pleistocene (880  
11 Kya, 95% posterior intervals 550 – 1,320 Kya) and estimate radiation of current northern  
12 hemisphere lineages between 50-200 Kya, indicating colonization of the northern oceans  
13 prior to the last glacial maximum. Coalescent analyses reveal restricted gene flow  
14 between ocean basins, with long-term migration rates (individual migrants per  
15 generation) of <3.3 for mtDNA and <2 for nuclear genomic DNA. Genetic evidence  
16 suggests that humpbacks in the North Pacific, North Atlantic and Southern Hemisphere  
17 are on independent evolutionary trajectories, supporting taxonomic revision of *M.*  
18 *novaeangliae* to three sub-species.

19

20

1 **1. INTRODUCTION**

2 The humpback whale (*Megaptera novaeangliae*) is an iconic and globally distributed  
3 migratory species. Within each ocean basin, humpbacks breed and calve in tropical and  
4 subtropical seas during winter, migrating to high latitudes to feed during summer. Despite  
5 an absence of geographical barriers to dispersal, populations show significant genetic  
6 structure between and within ocean basins (1-7) with the strongest restrictions to maternal  
7 gene flow across the equatorial boundary (1, 8). Observations of naturally marked and  
8 genotyped individuals suggest that maternally directed fidelity to both breeding and  
9 feeding grounds may be responsible for this population structure (9-14). Some population  
10 structure has also been identified in the nuclear genome; a recent study using multiple  
11 nuclear introns to survey Atlantic diversity (15) found evidence of population structuring  
12 between the North Atlantic and Southern Hemisphere, but not among breeding and  
13 feeding grounds within the North Atlantic.

14  
15 Although phylogenetic reconstructions of mtDNA show evidence of long-term gene flow  
16 between oceans (1), no permanent dispersal between populations in different hemispheres  
17 have been documented. Although seasonal breeding cycles are asynchronous between the  
18 hemispheres, two Southern Hemisphere breeding grounds extend north of the equator:  
19 Ecuador and Costa Rica in the Pacific (16) and Gabon and Guinea in the Atlantic (17,  
20 18), demonstrating that inter-hemisphere movements are biologically possible.  
21 Encounters on common breeding grounds between whales at the end or start of their  
22 respective winter breeding seasons could result in male-mediated gene flow, but genetic  
23 patterns of population structure and haplotype distribution show no evidence of this to  
24 date (7).

25  
26 Despite the evidence for limited gene flow between oceans, there has been no recent  
27 taxonomic investigation of humpback whales (19). In 1946, Tomilin (20) proposed  
28 humpbacks in the two hemispheres as subspecies, on the basis of a greater measured  
29 body length in the Southern Hemisphere form. Subsequent investigation (21) found no  
30 significant variation in lengths between oceans and, along with a later review of cetacean  
31 taxonomy (22), concluded that there was insufficient evidence for subspecies. Multiple

1 lines of evidence for genetic divergence (22) or the diagnosis of fixed character  
2 differences (23) could be a reason to revisit their status.

3  
4 Unlinked nuclear DNA markers provide multiple independent lines of evidence for  
5 reconstructing evolutionary histories of population structuring within species (24). Here  
6 we assess the genetic evidence for humpback global population structure, using the  
7 largest global genetic dataset for this species to date, including eight nuclear loci  
8 (~3,700bp in total length) from more than 70 individuals and mtDNA control regions  
9 from more than 2,900 individuals inhabiting all three ocean basins. We use conventional  
10 frequency and coalescent based population genetic approaches to (i) describe the pattern  
11 and magnitude of organismal (mtDNA) and gametic (nuclear DNA) gene flow between  
12 oceans, and (ii) estimate the time frame of radiation of extant humpback whale lineages.  
13 We consider these patterns of gene flow in the context of the criteria currently recognized  
14 as defining sub-species for cetaceans.

## 15 16 **2. METHODS**

### 17 **(a) mtDNA and nuclear datasets**

18 Sequences spanning 465bp of the mtDNA control region were compiled from North  
19 Pacific (5, 25, n=396), South Pacific and south-eastern Indian Ocean (13, n=804), south-  
20 western Atlantic (26, n=48), south-western Indian Ocean (27, n=1,137) and western  
21 North Atlantic (25, 28, n=348) studies. Blue (*Balaenoptera musculus*) and fin (*B.*  
22 *physalus*) whales were used as outgroups in phylogenetic reconstructions (GenBank  
23 AY582748, NC\_001321, NC\_001601 and AY582748) following (29). Sequences were  
24 aligned by eye in MacClade v4.0 (30). Another mtDNA dataset was assembled from  
25 shorter sequences, allowing greater sample representation from North Atlantic locations  
26 (3, 4, n=246); on alignment these overlapped with the 465bp worldwide dataset by  
27 285bp. Eight nuclear loci (*ACT*, *CAT*, *CHRNA1*, *ESD*, *FGG*, *GBA*, *LAC* and *RHO*;  
28 GenBank GQ407914-408186) were phased as described in Jackson *et al.* (25) for 70-80  
29 humpback whales worldwide (Figure 1). Exonic regions were excluded from analysis.  
30 ModelTest v3.7 (31) was used to determine the Akaike Information Criterion best fitting  
31 model for all datasets, with branch lengths included as parameters (32).

1

## 2 **(b) Mitochondrial population structure and gene flow**

3 Nucleotide diversity ( $\pi$ ) and haplotype diversity ( $h$ ) were estimated following Nei (33)  
4 using Arlequin v3.1 (34). Differentiation between oceans was estimated using  $F_{ST}$  and  $\phi_{ST}$   
5 with 50,000 matrix permutations. To correct for multiple substitutions,  $\phi_{ST}$  was adjusted  
6 using the Kimura 2-parameter +  $\Gamma$  mutation model (as supported by ModelTest for the  
7 humpback-only dataset). Hierarchical AMOVA tests were conducted to measure  
8 partitioning of variance (1) between oceans, (2) among regions (breeding and feeding  
9 grounds) within oceans, and (3) within regions (Figure 1). Two groupings were  
10 considered; (1) '5-oceans' (North Pacific, South Pacific, North Atlantic, South Atlantic,  
11 Southern Indian Ocean) and (2) three ocean basins ('3-oceans': North Pacific, North  
12 Atlantic and Southern Hemisphere).

13

14 The effective population size  $\Theta$  and numbers of effective female migrants per generation  
15  $2N_f m_f$  (equivalent to  $N_e m_f$ ) were estimated for both mtDNA datasets, partitioned by 3-  
16 oceans and 5-oceans, using the Bayesian inference program MIGRATE-N v3.5.1 (35,  
17 36). Each oceanic partition was sub-sampled to generate computationally tractable  
18 datasets containing 150 and 200 animals. Sampling was stratified within each ocean basin  
19 or ocean so that roughly equal numbers of samples were randomly selected from each  
20 breeding or feeding ground. Analyses were run with four Markov chains and gamma  
21 distributed priors on  $2N_f m_f$  (range 0-20 for 3 oceans, 0-100 for 5 oceans) and  $\Theta$  (range 0-  
22 0.1). The heating scheme was set to temperatures 1.0, 1.5, 3.0 and 100,000.0. Analyses  
23 were conducted with 100 replicates, with 10,000 steps recorded every 100 generations  
24 and 50% burn-in, totalling 50 million retained parameter values.

25

## 26 **(c) Mitochondrial phylogeny and divergence times**

27 A phylogeny of mtDNA control region sequences was reconstructed in Mr Bayes v4.0  
28 (37) using a HKY+ $\Gamma$  model of sequence evolution (as supported by ModelTest for  
29 humpback whales plus outgroups). Analysis was conducted for 25 million generations  
30 (sampling every 1,000 generations), with 10% discarded as burn-in and split frequencies  
31 monitored for convergence. Posterior parameter distributions were examined using

1 TRACER v1.5 (38).

2

3 To estimate within-species divergence times, a humpback-only dataset was analysed in  
4 BEAST v1.7.4 (39), containing 150 individuals from each of the 5 oceans, chosen by  
5 stratified random sub-sampling as above. Since humpback whales exhibit complex sub-  
6 structure, we tested the fit of three coalescent models: expansion, logistic and constant  
7 size, and to strict and relaxed lognormal clock rates. Analyses were run for 25 million  
8 generations. Bayes Factors were calculated in TRACER to determine the best fitting  
9 population model. We imposed a strict clock with a rate of 3.94% bp<sup>-1</sup> million years  
10 (MY)<sup>-1</sup> for the humpback control region as determined by phylogenetic approaches (25).  
11 To assess the impact of assumed substitution rate we repeated the analysis using a higher  
12 rate of substitution (mean 14.9% bp<sup>-1</sup> MY<sup>-1</sup>), as estimated by Ho *et al.* (40) for bowhead  
13 whales using ancient DNA sequences.

14

#### 15 **(d) Neutrality and population expansion**

16 Tajima's *D* (41, 42) and Fu's *F<sub>s</sub>* (43) were estimated to test for selection (versus  
17 population neutrality) worldwide, and for haplotype clades within each ocean basin.  
18 Mismatch distributions were generated to test null hypotheses of population expansion  
19 for the Southern Ocean basin and for the three Northern Hemisphere mtDNA clades  
20 using Arlequin (34), with 1,000 bootstrap replicates.

21

#### 22 **(e) Nuclear diversity, structuring and gene flow**

23 Nuclear heterozygosity was estimated for phased alleles and sequences ( $\pi$ ), and.  $F_{ST}$  and  
24  $\phi_{ST}$  estimates of differentiation between oceans and ocean basins were measured with  
25 50,000 permutations in Arlequin. We used Jombart's Discriminant Analysis of Principal  
26 Components (DAPC, 44) in the R package *adegenet* to evaluate the level of support for  
27 different numbers of distinct genetic clusters ( $K=1$  to  $K=20$ ) in the absence of *a priori*  
28 ocean divisions. DAPC can discriminate complex patterns like hierarchical clustering or  
29 stepping stone structures- realistic possibilities since humpbacks exhibit fidelity to  
30 multiple migratory routes between various breeding and feeding areas. Sequential *K*-  
31 mean clustering was applied to find the best-supported cluster size. All 21 principal

1 components (PCs) were retained initially. We then applied the discriminant function  
2 *dapc*, which constructs synthetic variables in order to maximise variation between, and  
3 minimise variation within, each cluster group. After inspecting the cumulative variance  
4 retained by the principal components, the *a*-score was used to calculate that 5 PCs are  
5 sufficient to characterise population structure. Based on these discriminant functions,  
6 posterior probabilities of membership to each of the three ocean basins (North Atlantic,  
7 North Pacific and Southern Hemisphere) were calculated for each individual and within  
8 each ocean basin.

9  
10 Population differentiation and partitioning of allelic variance was calculated: (1) within  
11 individuals, (2) among individuals within three ocean basins, and (3) among individuals  
12 between three ocean basins, using the standard AMOVA test in Arlequin (45). For this  
13 we included only individuals for which >75% of intronic loci had been sequenced.  
14 Effective population size for each ocean basin and effective numbers of migrants were  
15 co-estimated using MIGRATE-N (35, 36) with gamma distributed priors ranging  
16 between 0-0.1 and 0-20, respectively. Analyses were conducted for 100 replicates, with  
17 10,000 steps recorded every 1,000 generations and 50% removed as burn-in, totalling 500  
18 million parameter values retained.

19  
20 Extended nuclear DNA sequences (including intronic and exonic sequences described in  
21 25) were aligned with a selection of Balaenopteridae (blue, fin and Antarctic minke  
22 whales, *B. bonaerensis*) in MacClade (30). Patterns of allelic divergence between  
23 humpback, blue and fin whales were calculated using statistical parsimony networks with  
24 program TCS v1.21 (46).

### 25 26 **3. RESULTS**

27 The mtDNA control region sequences (length 465bp) were compiled for 2,733  
28 individuals worldwide (2,979 individuals at length 285bp). This corresponds to 1.3 mega-  
29 bases (Mb) of data. The eight nuclear loci corresponded to 3.7 kilo-bases (Kb), yielding  
30 140-160 alleles per locus, a total dataset size of 663Kb (Table 1).

31

## 1 (a) Mitochondrial diversity and phylogeny

2 Within the 465bp mtDNA control region dataset we found 85 variable sites, resolving  
3 219 haplotypes worldwide. The 285bp dataset contained 78 polymorphic sites, resolving  
4 209 haplotypes. In both cases, a number of sites have undergone multiple substitutions  
5 ('hits'). At these mutational hotspots, phylogenetic signal may be diminished by the noise  
6 created by multiple mutations (including back-mutations). Nearly all haplotypes in the  
7 465bp dataset were private to an ocean basin, with only one haplotype shared between  
8 the North Atlantic and Southern Hemisphere and two shared between the North Pacific  
9 and Southern Hemisphere. Control region sequences showed no fixed 'diagnostic'  
10 substitutions unique to each ocean basin.

11  
12 Total worldwide nucleotide diversity  $\pi$  was 2.14% for the 465bp dataset. Ocean basin  
13 estimates of  $\pi$  were similar to those obtained in previous studies (1, 7): 1.13% in the  
14 North Pacific, 1.97% in the North Atlantic and 2.48% in the Southern Hemisphere  
15 (Supplementary Tables 1, 2). The 285bp control region dataset yielded higher global  $\pi$   
16 (4.16%) because most variable sites in the 465bp dataset fall within the 285bp fragment.  
17 A similar pattern was observed for oceanic population size  $\Theta$ , which was similar for both  
18 northern oceans at 285bp consensus length (0.009) but lower for the North Atlantic at the  
19 465bp length (0.004) compared to the North Pacific (0.007), probably as a consequence  
20 of more limited geographic sampling of the 465bp dataset.

21  
22 The Bayesian majority-rule phylogeny (Ts:Tv=43.4,  $\alpha = 0.136$ ) of the 465bp haplotypes  
23 supported the grouping of humpback mtDNA control region sequences into 4 previously  
24 recognised clades (Supplementary Figure 1; 1, 13). The largest clade is 'CD' (96%  
25 Bayesian posterior probability support, PP), which contains haplotypes from all oceans  
26 and includes a haplotype shared across the Pacific equator. The next largest ('IJ', 82%  
27 PP) includes haplotypes from all oceans except the north Pacific, and one haplotype  
28 shared across the Atlantic equator. The smallest clade, 'SH' (89% PP) includes only  
29 Southern Hemisphere haplotypes. A fourth 'AE' clade contains mostly North Pacific  
30 haplotypes, but fell as a basal polytomy. The 285bp dataset expands the haplotypic  
31 diversity of the North Atlantic 'IJ' clades (Table 1) and increases support for 'IJ' (100%

1 PP), while providing weak support for clade CD (72% PP), and none for SH and AE,  
2 likely due to a reduction in variable sites (Supplementary Figure 2). The phylogenetic  
3 interrelationships among clades are not consistent and weakly supported. This may be  
4 due to saturation of the control region obscuring true signal, or rapid radiation of clades  
5 close to the origin of the present day humpback mtDNA lineages.

6  
7 Bayes factors (BF) supported the constant size model over other growth models ( $BF > 4$ ).  
8 Using the phylogenetically derived substitution rate (Figure 2), the median root age was  
9 880 Kya (95% PI 55-132 Kya). Northern ocean clades diverged from the Southern  
10 Hemisphere subsequently, with the earliest North Pacific clade radiating 170 Kya (95%  
11 PI 14-80 Kya) followed by a second radiation 70 Kya. The earliest North Atlantic clade  
12 radiated 87 Kya (95% PI 1.200-146 Kya), followed by subsequent radiation of two extant  
13 clades c. 55 Kya and 38 Kya. As the imposed molecular clock did not include any  
14 variance, however, these values provide only broad guidance rather than representing the  
15 full range of uncertainty. Using the ancient DNA-derived bowhead rate, the median root  
16 age was 232 Kya (95% PI 137-346 Kya) with the earliest northern ocean radiation (AE  
17 clade, North Pacific) dated to 47 Kya (95% PI 21-90 Kya) and the North Atlantic  
18 radiations ranging from 14-23 Kya (data not shown).

#### 19 20 **(b) Neutrality and population expansion**

21 The null hypothesis of population equilibrium was rejected by Fu's  $F_s$  for the Southern  
22 Hemisphere and North Atlantic 'IJ' haplotype clade (Supplementary Table 4), suggesting  
23 that both populations expanded in the past. For the Southern Hemisphere the SSD test for  
24 spatial (but not sudden) expansion was rejected, suggesting humpbacks may have  
25 undergone a sudden expansion in this ocean. For the 'IJ' clade, the test for sudden  
26 expansion was rejected, so this clade may have instead undergone a spatial expansion.  
27 Alternatively, true history may have involved more complex expansion scenarios not  
28 captured by these tests. Spatial and sudden population expansions were rejected for the  
29 North Atlantic 'CD' clade, which is estimated to diverge into the North Atlantic more  
30 recently than the 'IJ' clade. Test statistics do not support long-term expansion of any  
31 North Pacific clades.

1

2 **(c) Mitochondrial oceanic differentiation and gene flow**

3 The hierarchical AMOVA showed strong differentiation among oceans (Supplementary  
4 Table 4). Greater differentiation was found between the three ocean basins (28% and  
5 10% of total molecular and haplotypic genetic variation respectively) than between five  
6 oceans (18% molecular, 6% haplotypic), suggesting that gene flow has been more  
7 restricted between inter-hemispheric oceans than across the Southern Hemisphere oceans  
8 (Supplementary Table 5). Molecular differentiation was greater than haplotypic  
9 differentiation in both cases, indicating that substantial genetic divergence as well as drift  
10 has been occurring between the three ocean basins (47). The level of overall molecular  
11 differentiation between five oceans was similar to that between individual breeding and  
12 feeding populations (around 18% of total variation), suggesting similar genetic  
13 divergence at both spatial scales, and in both cases much lower divergence than that seen  
14 at the inter-basin scale.

15

16 The greatest population differentiation was found between the Northern Hemisphere  
17 oceans (mtDNA 465bp  $F_{ST} = 0.18$ ,  $\phi_{ST} = 0.51$  for 465bp mtDNA). Similar differentiation  
18 was found between the North Pacific and the three Southern Hemisphere oceans (all  $\phi_{ST} \approx$   
19 0.35; Table 2, Supplementary Table 6). Nucleotide differentiation of roughly half this  
20 magnitude was estimated between the North Atlantic and the three Southern Hemisphere  
21 oceans ( $\phi_{ST} = 0.16-0.18$ ). This may be because the North Pacific clade 'AE' diverged  
22 from the Southern Hemisphere earlier than the North Atlantic clades (Figure 2).  
23 Differentiation among oceans of the Southern Hemisphere was two orders of magnitude  
24 weaker (Supplementary Table 6), with varying levels of differentiation between Southern  
25 Hemisphere oceans (Rosenbaum et al. submitted).

26

27 Coalescent estimates of maternal gene flow between ocean basins were low (Figure 3).  
28 Gene flow (immigrants per generation) was slightly higher from the Northern  
29 Hemisphere to the Southern Hemisphere ( $\sim 3 N_e m_f$ ), with wider confidence intervals and  
30 an upper 95% boundary up to 8.5, compared to Southern Hemisphere movements into the  
31 North, with mean values of 0.6 – 1.1  $N_e m_f$  and an upper 95% boundary of  $< 2.8$  (Table 3).

1 The 5-ocean analysis showed a similar pattern of restricted gene flow across the equator,  
2 but gene flow between the Southern Hemisphere oceans (Supplementary Table 6) was so  
3 high that  $N_e m_f$  values were truncated by the maximum upper boundary of the prior,  
4 indicating little restriction to migration flow.

#### 6 **(d) Nuclear diversity and networks**

7 The numbers of alleles at each intron ranged from 2 (*GBA*) to 10 (*RHO*). A total of 50  
8 variable sites were identified across the entire dataset, of which six were  
9 insertions/deletions. Heterozygosity varied from 0.01 (*LAC*) to 0.22 (*RHO*). Within-  
10 ocean nucleotide diversity ( $\pi$ ) ranged thirty-fold across nuclear loci, from 0.00% (*LAC*) to  
11 1.32% (*RHO*). Average worldwide genomic  $\pi$  was 0.22%. Within ocean basins, mean  $\pi$   
12 diversity was 0.09% in the North Pacific, 0.16% in the North Atlantic and 0.12% in the  
13 Southern Hemisphere (Table 1).

14  
15 Most common alleles were found in similar frequencies among the three oceans  
16 (Supplementary Figure 3). A number of region-specific ('private') alleles were found in  
17 the Southern Hemisphere but there were no fixed or diagnostic differences. One highly  
18 divergent *actin* allele (48) is widely distributed and relatively common. This allele is  
19 equidistant between fin and humpback whale clades (average distance to these is 0.012-  
20 0.013), while average distance within the humpback clade is 0.006. The allele could  
21 represent an 'ancestral' balaenopterid lineage which originated prior to the evolutionary  
22 radiation of humpbacks and which might be under selection, considering the absence of  
23 closely related alleles. Alternatively the allele could be divergent due to past genetic  
24 introgression from other balaenopterids, e.g., by hybridization. *CAT* and *ESD* were  
25 strongly differentiated from nearest neighbours (8 and 19 mutation steps respectively  
26 from humpback whales). For other loci, the distance to outgroups was less than or equal  
27 to the maximum distance between alleles within humpbacks. Divergences among  
28 balaenopterids are therefore low for most loci, reflecting a slow mutation rate (25) and  
29 possibly also inter-species introgression (e.g. 49).

#### 31 **(e) Nuclear oceanic differentiation**

1 A weaker pattern of oceanic differentiation was seen in the nuclear dataset, compared to  
2 mtDNA (Supplementary Tables 8 and 9), with overall  $F_{ST}=0.12$  between the 3 ocean  
3 basins. When the Southern Hemisphere was partitioned into south-eastern Indian Ocean  
4 and South Pacific regions, this reduced to  $F_{ST}=0.06$ . Levels of differentiation between  
5 Northern Hemisphere oceans (combined nuclear  $F_{ST} = 0.15$ ) were similar in magnitude to  
6 those obtained for mtDNA, while differentiation between the two northern oceans and  
7 Southern Hemisphere was much weaker (combined nuclear differentiation from North  
8 Pacific and North Atlantic was  $F_{ST} = 0.05$  and  $0.09$  respectively, Table 2). No significant  
9  $F_{ST}$  or  $\phi_{ST}$  differentiation between the south-eastern Indian and South Pacific oceans was  
10 detected by any nuclear loci.

11  
12 DAPC analyses yielded 6-9 clusters as a good fit to the dataset. In each repeat analysis  
13 over  $K=6-9$ , a Southern Hemisphere-only cluster and predominantly North Atlantic  
14 cluster were stably recovered. Probabilities of membership within each ocean were all  
15 over 70% when 5 PCs were used (Supplementary Figures 4 and 5).

16  
17 Coalescent estimates of  $\Theta$  across loci (Supplementary Table 10) revealed a pattern of  
18 lower diversity in the Northern Hemispheres and higher diversity in the Southern  
19 Hemisphere. Coalescent migration rates between ocean basins (Figure 3) were slightly  
20 lower than those obtained from mtDNA, but in a very similar magnitude range, with  
21 upper percentiles  $<4$  migrants per generation. Similarly to mtDNA, gene flow into the  
22 Southern Hemisphere was greater than gene flow into the North Atlantic, but unlike  
23 mtDNA there was fairly symmetrical nuclear gene flow estimated between the North  
24 Pacific and Southern Hemisphere.

25

26

## 27 **4. DISCUSSION**

### 28 *Genetic diversity- cultural maintenance?*

29 Our diversity metrics reveal higher mtDNA nucleotide genetic diversity in humpbacks  
30 than other baleen whales (50-54), with comparable levels only found in the southern right  
31 whale (55). High nucleotide diversity may reflect large ancestral population sizes (53), or

1 may be driven by strong population structuring and restricted gene flow between  
2 populations. For humpback whales and southern right whales (55), mtDNA haplotype  
3 frequencies show marked differences between breeding/calving grounds. Photo-  
4 identification and genetic evidence suggests that this is driven by maternal fidelity to  
5 natal breeding grounds (e.g., 12), so this behaviour may be the major driver influencing  
6 high global mtDNA diversity levels. In contrast, nuclear genetic diversity of humpbacks  
7 by ocean basin is lower than comparable estimates in Antarctic minke whales (15), but  
8 similar to levels estimated for gray whales (54), suggesting that humpbacks and gray  
9 whales may have had smaller past population sizes or a greater loss of diversity as a  
10 consequence of population bottlenecks due to whaling.

11

#### 12 *Gene flow of baleen whales across the equator*

13 Strongly significant mtDNA differentiation of humpbacks in each of the world's ocean  
14 basins indicates that extensive genetic drift and mutational divergence has occurred  
15 between populations. However much more divergence has occurred between inter-  
16 hemispheric ocean basins, suggesting that each basin is isolated by equatorial barriers to  
17 movement. Our inter-population divergence levels are consistent with previous analyses  
18 finding significant differentiation and sometimes also divergence between breeding  
19 populations (13, 27), but here we demonstrate that humpback divergence between ocean  
20 basins is an order of magnitude greater, strong enough even to drive population  
21 differentiation in slowly evolving nuclear intronic genes. Recent worldwide analyses of  
22 fin whale mitogenomes also showed strong population divergence between the North  
23 Pacific, North Atlantic and Southern Ocean, suggesting similar restrictions to trans-  
24 equatorial gene flow for fin whales (56). Levels of mutational divergence ( $\phi_{ST}$ ), between  
25 the North Pacific and North Atlantic humpbacks are equivalent to divergence between  
26 right whale species inhabiting the Southern Hemisphere and North Atlantic ocean basins  
27 (51). However in contrast with right whales, no diagnostic mtDNA or nuclear sites have  
28 been identified between the two Northern Hemisphere oceans in this study for humpback  
29 whales.

30

1 Two types of gene flow have been measured in this study– gametic (from nuclear DNA)  
2 and female-mediated organismal (from mtDNA). If gene flow occurs as a result of  
3 whales from the two hemispheres mating at the extreme edge of their wintering seasons,  
4 such exchange would only be detected using nuclear genes since females remain in their  
5 natal hemispheres. Estimates of gene flow from the Southern to the Northern Hemisphere  
6 are similar for both nuclear and mtDNA (<1.6 migrants per generation), suggesting that  
7 both organismal and gametic exchange is infrequent and is not sex-biased (*e.g.*,  $R \sim 1$ , 57).  
8 The low migration rates reported here could be a consequence of regular, low level  
9 genetic exchange, or no regular exchange but occasional pulses of migrants over time,  
10 possibly as a consequence of unusual oceanographic or environmental conditions.  
11 Surprisingly, female-mediated gene flow is slightly higher than biparental gene flow  
12 across the Atlantic equator. Females crossing the equator are more likely to produce  
13 offspring than males (since males compete for mates), so this may be the driving  
14 mechanism. This suggests migration across the equator may have been more influential  
15 in determining Atlantic gene flow than mating on common wintering grounds (which  
16 would not be reflected in mtDNA gene flow).

17

18 In all cases, southward migration across the equator was higher, though not significantly  
19 so, than northward migration. Oceanic shifts in temperature shifted upwelling centres  
20 during periods of glacial expansion, which may have reduced available habitat in many  
21 Northern Hemisphere areas (*e.g.*, 58, 59). This reduction may have led to southward  
22 shifts in humpback distribution both on feeding grounds and possibly also breeding  
23 grounds, increasing the chance of southward gene flow.

24

25 *Oceanic subspecies?*

26 Reeves *et al.* (22) recommended that the ranking of subspecies be used to “embrace  
27 groups of organisms that appear to have been on independent evolutionary trajectories  
28 (with minor continuing gene flow), as demonstrated by morphological evidence *or* at  
29 least one line of appropriate genetic evidence”. We consider that oceanic populations of  
30 humpback whales meet these criteria. Two lines of genetic evidence support an  
31 independent evolutionary trajectory for humpback whales in the three ocean basins: 1)

1 differentiation and divergence of mtDNA, reflecting low organismal gene flow; and 2)  
2 differentiation of multiple nuclear DNA loci, reflecting reproductive isolation.

3  
4 (1) Mitochondrial DNA control region data shows strong divergence between ocean  
5 basins, with only 3 haplotypes shared between the ocean basins in the 465bp dataset.  
6 Although there are no diagnostic sites, nor reciprocal monophyly of haplotypes between  
7 ocean basins, coalescent-based measurements of inter-oceanic gene flow by females are  
8 <1 migrant per generation between some oceans with a maximum of <4 migrants per  
9 generation.

10  
11 (2) Nuclear DNA shows evidence of differentiation of allele frequencies and mutational  
12 divergence, although no diagnostic differences between ocean basins are present. The  
13 latter is unsurprising considering the slow rates of mutation estimated for these loci  
14 (0.05%/MY, 25). However nuclear loci can be used to assign individuals to ocean basins  
15 (>70%), and estimates of nuclear (bi-parental) gene flow are <1 whale per generation  
16 between some oceans and a maximum of <2 migrant whales per generation in all  
17 comparisons, despite the relatively slower rate of genomic drift compared to mtDNA.  
18 These low rates suggest that populations in different ocean basins have been  
19 reproductively isolated, as well as isolated by maternal traditions within oceans.

20  
21 Based on our results, and given the potential revision into oceanic subspecies, we propose  
22 the following names: *M. n. kuzira* (Gray, 1850) for the North Pacific, *M. n. novaeangliae*  
23 (Borowski, 1781) for the North Atlantic and *M. n. australis* (Lesson, 1828) for the  
24 Southern Hemisphere (60).

#### 25 26 *Pleistocene divergence and expansion*

27 While the radiation of current worldwide humpback lineages lies within the Pleistocene,  
28 more precise dates remain uncertain. Phylogenetic substitution rates (25) place these  
29 divergences within the timeframe of the last million years, with colonisation of the  
30 northern oceans by modern mtDNA lineages within the last 200,000 years. Relative clade  
31 ages suggest that modern lineage divergence into the North Pacific came earliest- the

1 'AE' clade is estimated to radiate c. 175,000 years ago, but diverges over 500,000 years  
2 ago from other haplogroups. Multiple significant incursions are then made into both  
3 northern oceans in the last 100,000 years, once into the North Pacific and at least twice in  
4 the North Atlantic. Only in the North Atlantic 'IJ' clade is there evidence for northern  
5 population expansion; the data indicate a spatial (slow advance) expansion, rather than a  
6 rapid radiation, suggesting that this clade may have been the first to expand into the  
7 North Atlantic, possibly after retreating ice in the past. This is consistent with reports that  
8 the North Atlantic 'IJ' haplotype clade is more broadly distributed across the North  
9 Atlantic than the 'CD' clade (3, 7). Differences in female reproductive success may  
10 however also influence clade patterns in this ocean basin (e.g., 28).

11  
12 Despite the low mtDNA diversity of North Pacific humpbacks (1, 5), analyses of  
13 divergence times suggest an early split of the North Pacific 'AE' clade from other  
14 humpback lineages worldwide, estimating the earliest divergences within the 'AE' group  
15 >150,000 years ago. Population expansion metrics show no signs of a radiation, although  
16 none is rejected either. The low diversity of this clade may therefore be driven by non  
17 age-related factors. Prolonged periods at small population size may increase apparent  
18 population divergence due to genetic drift, although humpback whale substitution rates  
19 are low (25) so this effect would have to persist over many thousands of years. It has  
20 been suggested that whaling led to the reductions in genetic diversity currently observed  
21 in other matrilineal species such as right whales (55, 61) and bowheads (62). A recent  
22 bottleneck due to whaling, and/ or deeper historical factors (such as reduction of feeding  
23 areas during glacial periods) are possible explanations.

24  
25 There is a general debate over whether phylogenetic substitution rates are biased  
26 downwards when considering within-species radiations (e.g., 63). No calibrations within  
27 the species are available to test whether this is the case for humpback whales. Applying a  
28 within-species rate derived from bowhead whales via ancient DNA (40) yields much  
29 more recent divergence times among humpback clades. If such rates turn out to be more  
30 accurate, the timeframe of radiations would be much more recent, with for example the  
31 North Atlantic 'IJ' clade estimated to c. 55,000 years before present. A more

1 representative collection of mitogenomic sequences (e.g., 25) and additional estimates of  
2 population-level humpback mutation rates are required to resolve this uncertainty. Sea  
3 surface temperatures and ice sheet reconstructions of the last glacial maximum (LGM)  
4 suggest that in the North Atlantic the Norwegian Sea was reliably free of ice during the  
5 summer months (64) whereas the Gulf of Maine and Scotia Shelf regions show evidence  
6 of grounded ice reaching to the continental shelf edge during that period, c. 21-22,000  
7 years ago (58). Refugia in the eastern North Atlantic may therefore have been more  
8 extensive, although it is also possible that primary foraging areas were just shifted south  
9 during LGM periods. In the western North Pacific, cores from the Sea of Okhotsk suggest  
10 sea ice cover may have been perennial during the LGM (59), but data from areas to the  
11 east is patchy. Confidence intervals on our population expansion statistics are broad and  
12 do not exclude the possibility of post-LGM re-colonization, but considered in concert this  
13 evidence suggests *de novo* colonisation of the northern oceans by humpbacks after the  
14 LGM (>12,000 years ago) is unlikely and that humpback persistence in these regions has  
15 a much longer history.

16

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- 8  
9  
10

1 **Figure legends**

2

3 Figure 1. Worldwide distribution of humpback whale mtDNA source locations.  
4 Wintering regions (circles), feeding grounds (crosses) and migratory routes (gray  
5 circles) are shown. Double circles show nuclear DNA source locations. Double-  
6 lined boxes denote ocean-basin groupings.

7

8 Figure 2. Bayesian chronogram of divergence and radiation of humpback  
9 mtDNA haplogroups (465bp), identified as 'CD', 'AE', 'IJ' and 'SH' (following  
10 1, 13). North Atlantic clades are red, North Pacific clades are blue, and Southern  
11 Hemisphere clades are unshaded. Dashed lines show 95% posterior estimates of  
12 key divergence times.

13

14 Figure 3. Posterior distributions of migration rates ( $N_e m$ ) between ocean basins  
15 from MIGRATE. MtDNA posteriors in black (465bp solid, 285bp dashed)  
16 represent mtDNA gene flow  $N_e m_f$ . NuDNA posteriors (red) represent nuclear  
17 gene flow ( $N_e m_{f+m}$ ). Prior distribution = dashed gray.

1 Table 1. Basic diversity estimates of nuclear genomic and mtDNA control region sequences used in this study.

	<i>ACT</i>	<i>CHRNA1</i>	<i>GBA</i>	<i>CAT</i>	<i>LAC</i>	<i>RHO</i>	<i>ESD</i>	<i>FGG</i>	<i>Introns</i>	<i>CR 465</i>	<i>CR 285</i>
# chromosomes (2n)	550	158	158	140	146	156	140	142	170	2733	2979
Length (bp)	474	306	118	422	334	122	661	1008	3445	465	285
# polymorphic sites	11	3	1	3	3	5	11	13	50	85	78
# indels	1	0	0	0	0	2	2	1	6		
Alleles: North Atlantic	3(0)	2(0)	1(0)	2(0)	1(0)	10(4)	5(0)	4(1)		22(21)	41(38)
Alleles: North Pacific	4(1)	2(0)	2(1)	2(0)	2(1)	6(0)	5(1)	6(3)		19(17)	18(16)
Alleles: Southern hemisphere	7(3)	3(0)	1(0)	4(2)	3(2)	7(0)	7(2)	3(0)		181(178)	153(148)
Total # alleles	8	3	2	4	4	10	8	7		219	209
Obs Heterozygosity	0.2418	0.0971	0.0127	0.1762	0.0137	0.2237	0.1286	0.0607	0.1983	0.9846 <sup>a</sup>	0.9828 <sup>a</sup>
Standard deviation	0.2389	0.0073	0	0.2561	0	0.0790	0.1517	0.1256	0.1758	0.0006	0.0006
$\pi$	0.0061	0.0010	0.0001	0.0013	0.0000	0.0132	0.0023	0.0008	0.0022	0.0214	0.0416
SD	0.0035	0.0011	0.0006	0.0012	0.0002	0.0087	0.0015	0.0006	0.0012	0.0108	0.0208

- 2 DLP 465 and 285 refer to the two mtDNA datasets, 'Introns' shows statistics summed over all intronic loci. Numbers in parentheses  
3 represent alleles private to each ocean basin. Levels of nucleotide diversity ( $\pi$ ) and their standard deviations (S.D.) are reported. <sup>a</sup>  
4 Haplotype diversity is reported for the control region dataset.

1 Table 2. Inter-ocean genetic differentiation of humpback whales at nuclear loci and the 465bp mtDNA control region

	Southern Hemisphere		North Pacific		North Atlantic	
	mtDNA	nuDNA	mtDNA	nuDNA	mtDNA	nuDNA
SH	0.0248	0.0016	<b>0.3193</b>	<b>0.0451</b>	<b>0.1613</b>	<b>0.0990</b>
NP	<b>0.0858</b>	<b>0.0400</b>	0.0113	0.0009	<b>0.5164</b>	<b>0.1516</b>
NA	<b>0.0926</b>	<b>0.0610</b>	<b>0.1755</b>	<b>0.1030</b>	0.0197	0.0016

2  $F_{ST}$  and  $\phi_{ST}$  measures are shown below and above the diagonal respectively. Within-ocean diversity of each locus ( $\pi$ ) is shown in the  
 3 shaded diagonal (Tajima-Nei corrected pair-wise distances for introns, Kimura 2-Parameter correction for mtDNA control region  
 4 sequences,  $\alpha = 0.1364$ ). Bold text indicates values significant at  $p < 0.05$  after Bonferroni correction.

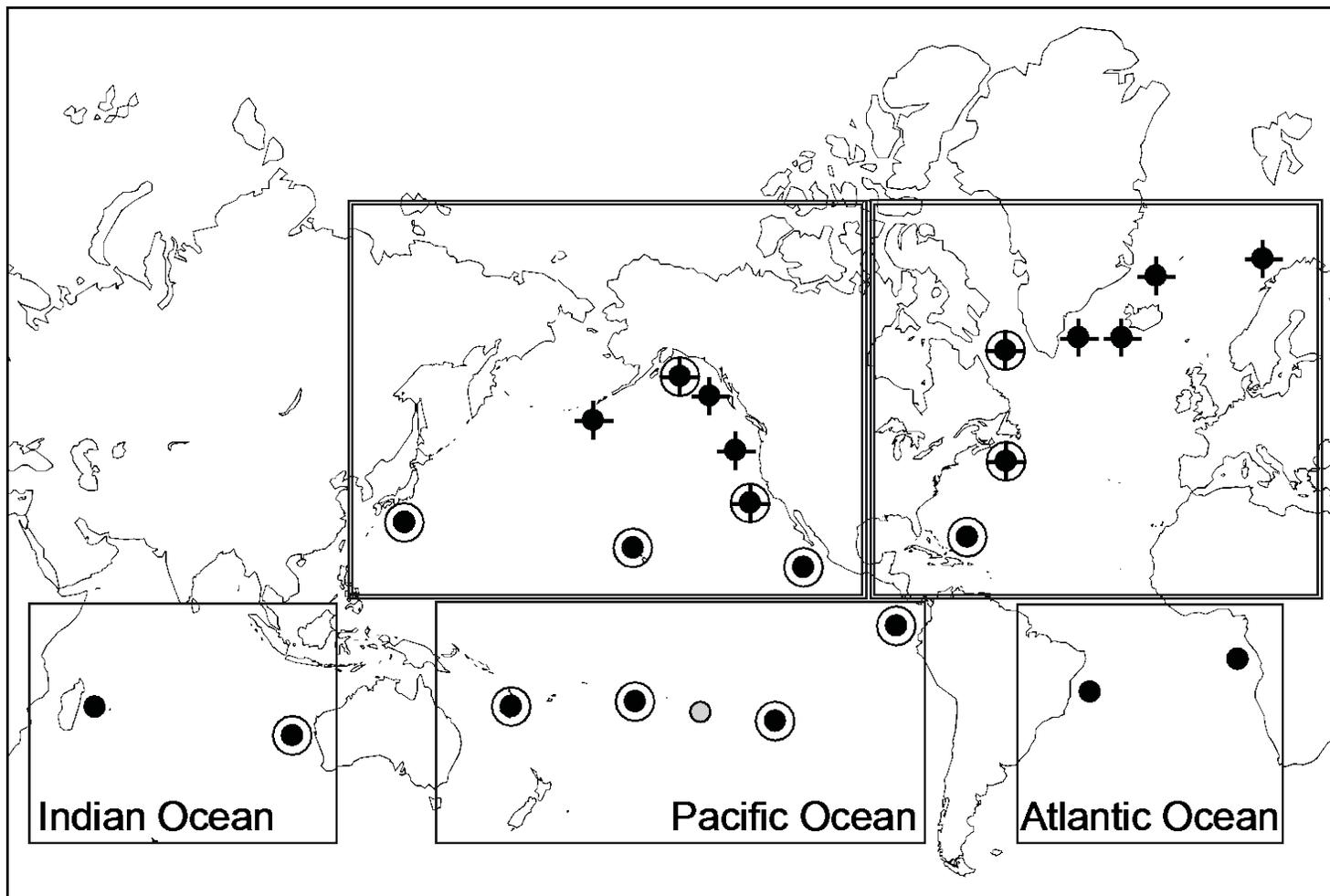
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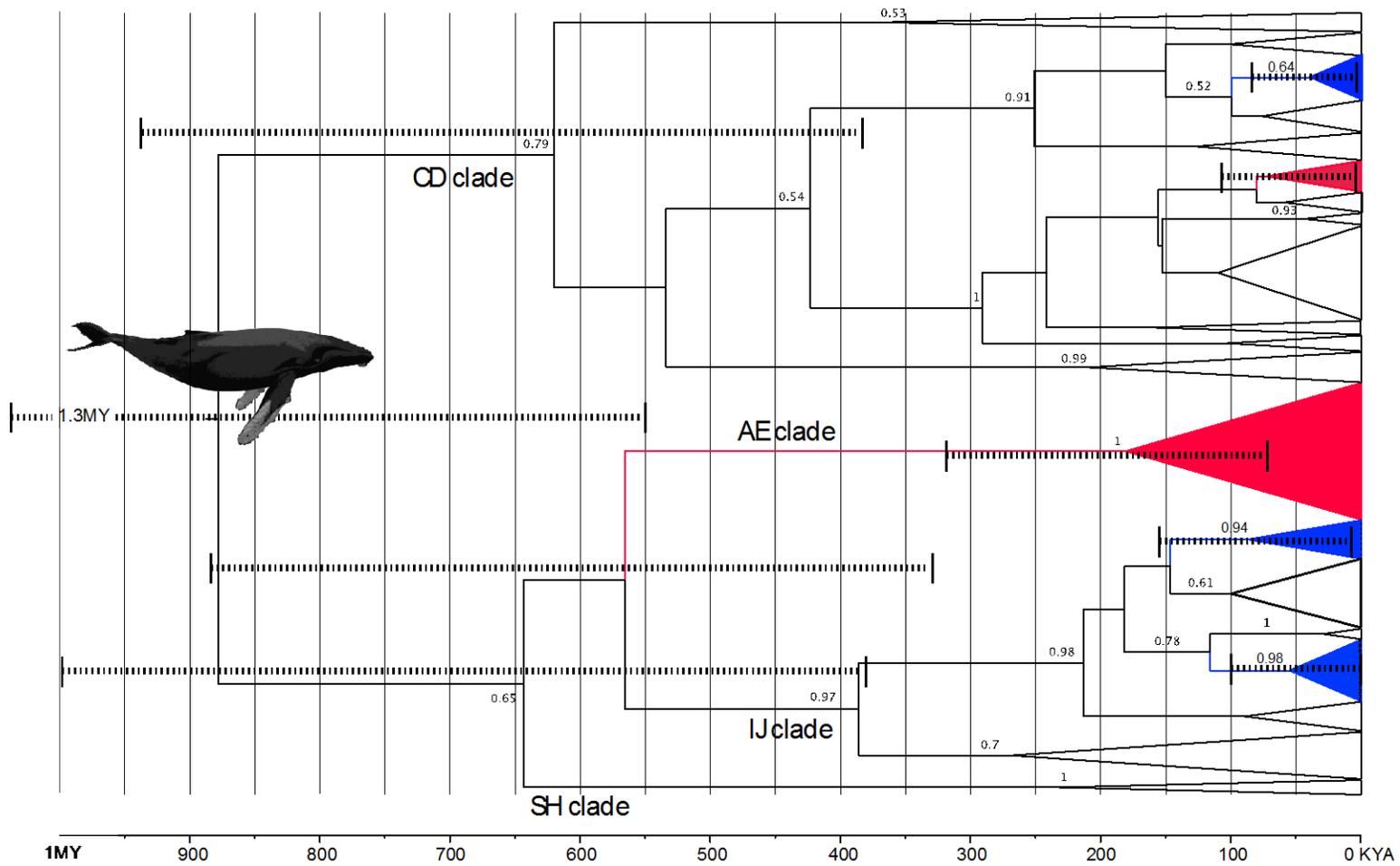
Table 3. MIGRATE  $N_{em}$  coalescent estimates of gene flow between ocean basins

		Southern hemisphere (SH)		North Atlantic (NA)		North Pacific (NP) 3	
		Mean	95% P.P.	Mean	95% P.P.	Mean	95% P.P.
465bp	From SH			0.87	0.00-2.08	0.69	0.00-1.85
	From NA	3.26	0.00-8.15			0.40	0.00-1.25
	From NP	2.98	0.00-8.28	0.26	0.00-0.85		
285bp	From SH			1.07	0.00-2.71	0.72	0.00-1.81
	From NA	3.87	0.00-9.20			0.33	0.00-1.03
	From NP	3.27	0.04-7.81	0.31	0.00-0.99		
nuDNA	From SH			0.82	0.00-2.08	1.51	0.00-3.59
	From NA	1.04	0.00-2.54			0.58	0.00-1.60
	From NP	1.65	0.00-3.84	0.92	0.00-2.40		

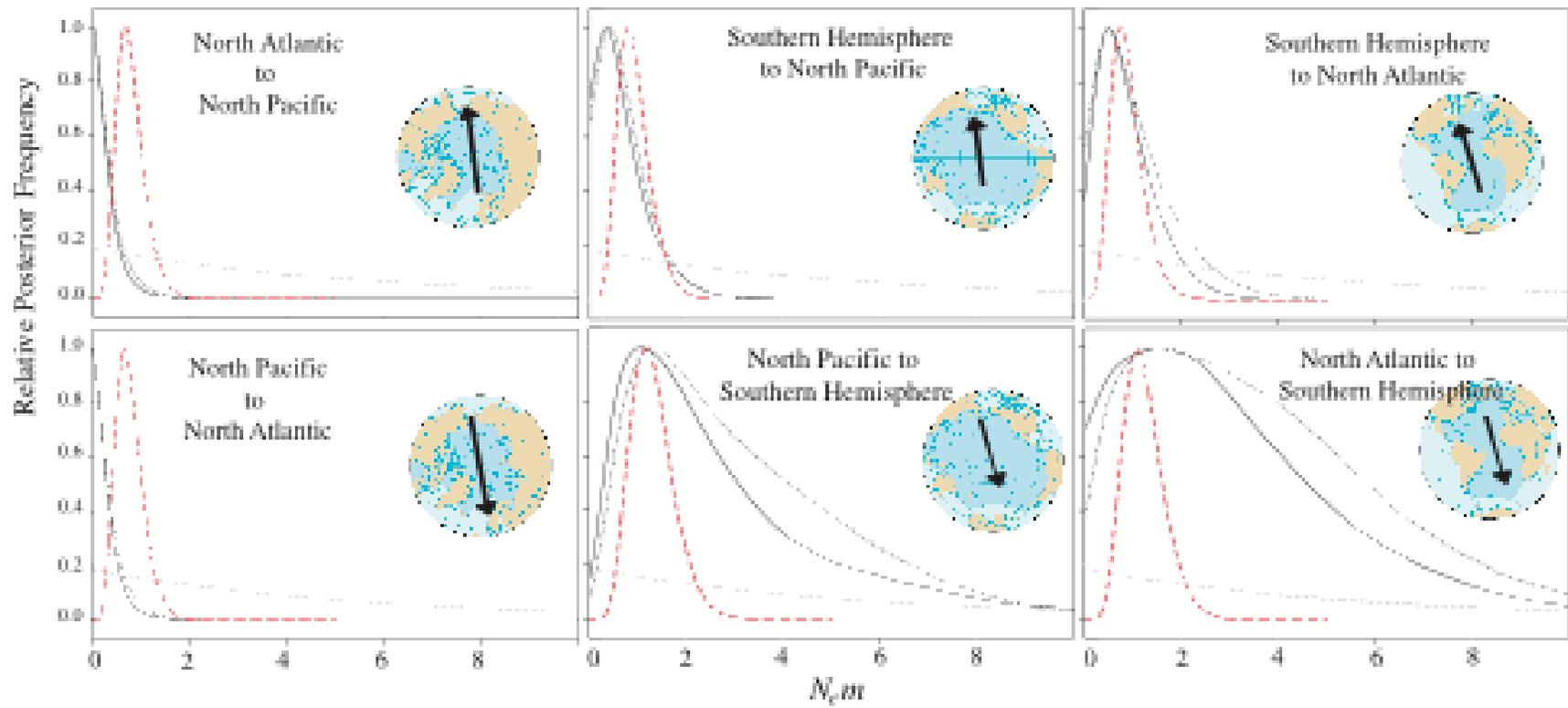
nuDNA ( $4N_{em_{f+m}}$ ) is divided 4 for comparability with mtDNA. P. P. refers to posterior probabilities.



1  
2 Figure 1.  
3



1  
2 Figure 2.  
3



1  
2 Figure 3.